



The environment recording unit in coral skeletons: structural and chemical evidences of a biochemically driven stepping-growth process in coral fibres

J. P. Cuif, Y. Dauphin

► To cite this version:

J. P. Cuif, Y. Dauphin. The environment recording unit in coral skeletons: structural and chemical evidences of a biochemically driven stepping-growth process in coral fibres. *Biogeosciences Discussions*, 2004, 1 (1), pp.625-658. hal-00330218

HAL Id: hal-00330218

<https://hal.science/hal-00330218>

Submitted on 29 Sep 2004

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Biogeosciences Discussions is the access reviewed discussion forum of *Biogeosciences*

The environment recording unit in coral skeletons: structural and chemical evidences of a biochemically driven stepping-growth process in coral fibres

J. P. Cuif¹ and Y. Dauphin²

¹Université Paris XI-Orsay, Bat. 504, Géologie, UMR IDES, F-91405 Orsay, France

²Université Paris VI-UPMC, Micropaléontologie, t. 46–56 E5, 4 place Jussieu, UMR IDESF, F-75252 Paris cedex 05, France

Received: 31 August 2004 – Accepted: 21 September 2004 – Published: 29 September 2004

Correspondence to: J. P. Cuif (cuif@geophy.geol.u-psud.fr)

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

Abstract

To improve our understanding of the environment recording by scleractinian corals, a detailed study of the skeleton microstructure has been carried out. A series of physico-chemical in situ characterizations was made, an approach that provides us with structural and biochemical information at the micrometric and nanometric scales. Gathering of these data results in a significant change in our concept of the growth of coral skeletons. In contrast to the usual view of an aggregate of purely mineral units (the coral fibres) independently growing by a simple chemical precipitation, coral skeletons appear to be biochemically controlled structures. Both structural and biochemical data reveal the micron-scaled stepping growth-mode of fibres, and its global coordination. In this process, sulfated acidic proteoglycans probably play a major role, due to their ability to create polymeric frameworks. Atomic force microscopy confirms the close relationship of organic and mineral phases at the nanometric scale.

A new microstructural model of coral skeleton formation is proposed, that places coral skeletons among the typical “matrix mediated structures”. From a practical standpoint, these results may contribute to develop a new high resolution approach in the study of paleoenvironments.

1. Introduction

Various chemical or isotopical proxies have placed coral skeletons among the most important sources of environmental information but, surprisingly, no agreement exists about the formation of these widely used biological archives. The process by which the basal ectoderm of coral polyps produces the underlying skeleton was the matter of a first controversy by the end of the 19th century when, in contrast to the von Heider's theory (1881), it was recognized that calcification does not occur within the ectoderm cells themselves. An extracellular calcification occurring outside the ectodermal cell layer, as advocated by von Koch (1882), gained general agreement. Since this time,

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

the control of skeleton organization remains an impending question. As summarized by Le Tissier (1991), models of calcification in corals range from a pure physicochemical (i.e. Barnes, 1970) to a biologically controlled process (i.e. Johnston, 1977–1980).

Crystallization of the coral aragonite being an extracellular phenomenon, the hypothesis of a biological control exerted on skeleton formation should be supported first by evidences concerning the place where crystallization occurs: the interface between the polyp basal ectodermal cell layer and the underlying skeleton. Noticeably, in spite of the Goreau's pioneering investigation (1956–1959), research focussing on physico-chemical characteristics of the subectodermal space are extremely rare. The complexity of coral skeletons prevents an easy access to the interface compartment, in contrast to Molluscs (i.e. Pelecypods), where the mineralization space between the outer side of the mantle and the shell is much more easily accessible. As a result, extraction of mineralizing fluids to study their composition and mineralizing capabilities was carried out in Molluscs as early as 1974–1976 (Wada and Fujinuki, 1974). To date, no equivalent research exists for coral mineralization process. The first pH measurement at the interface between the basal ectoderm and the growing surface of a coral skeleton was attempted in 2003 only (Al-Horani et al., 2003).

Instead of an unapproachable direct analysis of the mineralizing sites, complete decalcification of coral skeletons that give access to organic materials entrapped during skeletal growth has been much more used by investigators. This resulted in a series of papers that have emphasized the importance of either sugars (Wainwright, 1963; Wilfert and Peters, 1969; Dauphin, 2001) or amino acids (Young, 1971, 1973; Mitterer, 1978; Constantz and Weiner, 1988; Cuif et al., 1999). Nevertheless, as pointed out by Johnston (1980), research dealing with biochemical composition of organic materials extracted from coral skeletons results in mineralization models made “in ignorance of this material spatial distribution and micro architecture within the skeleton”.

Additionally, uncertainty persists with respect to the amount of these organic components. Since Wainwright (1963), the fraction of organic material embedded in the coral skeletons has been commonly estimated to 0.1%wt, a proportion recently reevaluated

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

I◀

▶I

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

to about 1%wt (Cohen and McConnaughey, 2003). On the other hand, thermogravimetric analyses (TGA) have shown that 2.5 to 3%wt of coral skeletons were lost before aragonite thermal decomposition (Cuif et al., 1997). Such a discrepancy casts doubts on the possible influence of these organic compounds on the crystallization process.

5 In response to the Johnston's remark, this paper aims to gather a series of recent data based on a different approach. Owing to the development of high resolution analytical instruments, combination of results from scanning electron microscopy, synchrotron X-ray fluorescence and atomic force imaging allows a fine scale in situ physico-chemical characterization of coral skeletons to be obtained. This results in
10 a new set of information about coral skeleton units examined at the micrometric and nanometric levels, allowing the calcification process to be reexamined.

2. Material and methods

Coral specimens have been collected alive, mostly by authors during the last few years. To remove tissues, polyps were decayed by a three-hour immersion in pure water that
15 causes the cells to be destroyed. Removal of the remaining living tissues was done by using water-jet, followed by an immersion in 0.1M sodium hypochlorite to clean the deeper parts of the skeletons. Then, specimens were rinsed and room-temperature air dried. In all cases, only the very upper parts of skeletons were used for imaging. The specimens were selected by SEM observation of polished and etched surfaces,
20 allowing the fibrous microstructures to be examined. Samples invaded by endolithic borers were discarded.

2.1. Materials

Corallites belonging to the following species have been used: *Favia stelligera* from Polynesian Archipelago (Moorea Island); *Montastrea curta* from Mururoa Island (J.P. Chevalier coll.); *Diploastrea heliophora*, *Leptoria phrygia*, *Porites* c.f. *australiensis*, *Acro-*
25

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

pora digitifera, *Merulina scabricula* from New Caledonia lagoon; *Caryophyllia ambrosia* from Atlantic Ocean (Gascogne Gulf); *Cladocora caespitosa* from Mediterranean Sea (Marseille coastal region); *Favia fragum* from Carribean Sea (Guadalupe Island).

2.2. Methods

5 2.2.1. Structural and chemical characterizations

Microstructural features of fibres have been first checked by microscope observation of ultra-thin sections in polarized light. This improvement of the usual microscopic technique requires fine polishing of the sample surface and a reduced thickness of slides. Instead of the standard 30 microns, grinding is continued up to some 4–5 micrometers thickness, that allows the primary colours of aragonite to be visible. Final polishing of the upper surface ensures an accurate microstructural observation.

2.2.2. Scanning Electron Microscopy (SEM) of coral skeleton

Observations were carried out not only on simple fracture surfaces but mostly on polished and etched surfaces. Etching by very light acidic solutions (one per mil formic acid including the fixative glutaraldehyde 3 to 5%) reveals that differences in solubility do exist within the crystal like fibres. Additionally, the global organization of coral septa is well evidenced, with emphasize on the trace of the early mineralization zone (commonly called “centres of calcification”).

2.2.3. X-ray Absorption Near Edge Spectrum (XANES) characterization and mapping of sulfated polysaccharides

At the European Synchrotron Radiation Facility in Grenoble (France), the ID-21 line has developed a high resolution analytical device based on detection of X-ray fluorescence. By carefully selecting X-ray wavelengths, characterization of bound energy in different oxidation state of sulfur is possible. After a calibration phase that allows

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

the different XANES spectra from sulfated standard molecules to be clearly separated (Figs. 1a to 1d), in situ characterization of organic sulfate can be made. Moreover, high performance lense focalization allows this characterization to be obtained with an infra-micronic spatial resolution. A two-dimensional piezoelectric driven specimen holder makes possible a micronic step displacement of the polished sample surface, resulting in a biochemical mapping of the coral skeleton in response to the selected energy.

2.2.4. Atomic Force Microscopy (AFM)

Scanning probe microscopy encompasses a family of techniques that measures surface topography and properties at the atomic scale. The atomic force microscope simultaneously produces maps the topography of surfaces (height images or derived amplitude images) and phase images. In phase imaging, a variant of tapping mode, the phase lag of the cantilever oscillation relative to the signal sent to the cantilver's piezo driver is used as a basis for image generation. Phase images can be generated as a consequence of variations in material properties such as composition, viscoelasticity, adhesion. AFM observations were conducted with Digital Instruments (Veeco) Nanoscope III Dimension 3100 at room temperature and air. The probe consisted of a cantilever with integrated Si_3N_4 tips (Digital Instruments). Micron scale images were acquired using tapping mode.

For AFM observation of biominerals, no routinely applicable procedure exists. Various preparative process of the observed surface have been used, that all aim to reduce the possible changes in relationships between mineral and postulated organic components. Consistency between phase and height images is essential for interpretation. The procedures of the sample preparations are given in the figure captions.

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

3. Results

3.1. The incremental growth mode of fibres and global microstructural patterns evidencing the stepping growth mode of coral skeletons

Checking of the microstructural characteristics emphasizes the common patterns of coral fibres: they appear as groups of elongated units (Fig. 2a) with global monocrystalline behaviour in polarized light (Figs. 2b, 2c). However, on a polished and etched surface, a completely different pattern appears, that reveals the incremental growth of fibres (Fig. 2d). This result is exemplified here by a microstructural sequence in a *Porites* c.f. *australiensis* colony that shows the morphology of skeletal component (Figs. 3a–3c). The fractured skeletal rod (Fig. 3c) shows the radial organization of fibres (Fig. 3d). Ultra-thin section (Fig. 3e) confirms the monocrystalline behaviour of fibres, whereas after etching the concentric growth lines transversal to fibre c-axes are well visible (Fig. 3f).

To date, no exception is known to this micron-scaled incremental growth of fibres. Attention must be drawn on the fact that visualisation of the stepping growth patterns depends on orientation of fibres with respect to the observation surface. It is long known that tridimensional arrangements of fibres are complex and very different among corals from various families. With respect to growth direction (Fig. 4a, white arrows) fibres are globally oblique to the septal median plan or growth axis. Consequently, within a given surface (Fig. 4b, line S), fibres may appear under different sections between two extreme conditions exemplified in Fig. 4c: from perfectly longitudinal (*L*) to transversal sections (*T*). Figures 4d and 4e give equivalent SEM pictures, showing that incremental growth patterns is variously readable depending on orientation of fibres.

Etched surfaces also show that fibres do not grow as independent units. Between adjacent fibres, a well-marked synchronism in the high solubility zones creates concentric lines evidencing the existence of a cyclic growth process. When the global orientation of fibres is favourable, continuity of concentric growth lines can be observed in the whole septum (Figs. 5a–5c), showing that this stepping growth pattern is coordi-

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

nated at a global scale. The series of few microns thick growth layers surround regions that exhibit specific microstructural patterns, in which Ca-carbonate shows a granular microstructure (Fig. 5c, EMZ). Specificity of these internal regions is also evidenced by the results of biochemical maps (see below 3.2).

- 5 3.2. Biochemical maps: layered distribution of sulfated polysaccharides within fibres, high concentration of S-polysaccharides in Early Mineralization Zones

All polished surfaces exposed to the 2.4825 keV X-ray energy produce a strong and exclusive signal corresponding to the presence of organic sulfated polysaccharides (Fig. 6). In contrast, sulfated aminoacids are practically undetectable. To take the
10 full advantage of the ID-21 mapping device, a preliminary exploration of the surface is useful, as exposed in Figs. 7a–7c. Part of a polished section (wall and septa) in a *Montastrea* corallite is firstly exposed to UV light (365 nm) on a reflection microscope, allowing the EMZ zones to be localized (Fig. 7c, arrows). The infra micronic 2.4825 keV beam, is then applied to the specimen, the distance between measurement points
15 and lines being one micron. Time exposure for each point is 0.8 second.

This results in the map of sulfur (Fig. 7d), i.e. the distribution of sulfated polysaccharides between EMZ and fibres on a 80×100 mm field view. Presently no precise quantitation is possible, owing to the heterogeneity of the biogenic material that does not allows any calculation concerning the absorbed/emitted radiations. However, the
20 higher concentration of polysaccharides in EMZ is well visible, as well as the banding patterns in the fibrous part of the field view (arrows).

Very comparable results are obtained on corallite from other species: *Diploria labyrinthica* (Figs. 8a–8d), *Porites* c.f. *australiensis* (Figs. 9a and 9b), *Acropora digitifera* (Figs. 9c and 9d). In all samples, a correspondence is evidenced between the
25 microstructural pictures and the biochemical maps. The exact correlation between the mineral growth layer arrangements and spatial distribution of acidic polysaccharides detected by their sulfated sulfur indicates that organic and mineral components are

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

interrelated within the growth layer itself, i.e. at an infra-micronic scale.

3.3. Spatial relationships between organic and mineral phases at the nanometric scale

5 AFM examination confirms the infra-micronic spatial relationships between organic and mineral components within skeleton growth layers. Pictures of *Merulina scabricula* are particularly significant. Densely packed nanograins are well visible (Figs. 10a and 10b). Looking more closely (Fig. 10c and 10d), the grains appear coated by a thin layer made of a material that exhibits strongly different physico-chemical properties, as shown by the phase image (Fig. 10d). Due to effectiveness of AFM phase imaging to
10 detect properties (i.e. elasticity) linked to chemical compositions, we can assess that the thin layers that appear as low relief at the grain surfaces (Fig. 10c, arrows) are very different in chemical composition from the grains themselves (Fig. 10d). Remarkably, similar pictures have been obtained on coral skeletons of every studied species, and at any place in the fibrous structures of each specimen (Figs. 11a–11b: *Favia stelligera*;
15 Fig. 11c–11d: *Cladocora caespitosa*; Fig. 11e–11f: *Caryophyllia ambrosia*).

This simultaneously shows that: (1) a nanometric granular structure is the basic organization of the fibre calcareous growth layers and (2) the few tenths to hundred nanometre grains are included in a material that exhibits different physico-chemical properties, suggesting an organic composition.

20 4. Discussion

When Johnston published his transmission electron microscope pictures (1977, Fig. 1; 1980, Fig. 18), the organic reticulum visible within the upper growth layer of a coral fibre was the first ever seen evidence of a sub micronic relationship between organic and mineral material within coral skeletons. Of course, no mineral was visible within
25 the reticulum spaces, due to the TEM preparative process.

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

4.1. Consistency of AFM data with Johnston results

Dimensions of the reticulum units in Johnston pictures remarkably correspond to those of the nanograins evidenced by AFM (Figs. 9–10). This correspondence fully supports the interpretation that the grains visible on the AFM pictures are made of aragonite.

- 5 Accordingly, the material that separates the mineral grains, with its strongly interactive properties (in phase images, black means “attractive”), can be supposed to be made of organic material.

Owing to the preparative process, Johnston was able to characterize the organic reticulum at the growing tips of fibres only. He suggested that the reticulum might
10 “disappear” in deeper parts of the skeleton. In contrast, we have noted that the XANES signal of sulfated polysaccharides is obtained on the whole surface of the sections, whatever the distance to the growing surface. Although the accurately focussed X-ray beam is less than 1 micron in diameter, it is much wider than the nanograin mean diameter. Therefore the polysaccharide signal is always observed, whatever the place
15 at which the X-ray beam is applied. Thus, the sulfated polysaccharide signal persists in coral skeletons far deeper than Johnston thought. However, there is no doubt that some change may occur within a coral colony lifetime, specifically in the global composition of organic compounds (Cuif et al., 1997).

Recent data show that the organic component in coral skeletons structurally includes
20 water (Cuif et al., 2004), a result that also contributes to create a consistent view of this material. From a quantitative stand point, the hydrated matrix of coral skeletons has been shown to represent up to 2.5 - 3% of the skeleton total weight, depending upon species. More than a half of this weight is made by water, a result that is close to the proportion suggested by Gaffey (1988) from infrared observations. Thus, biochemical
25 compounds (properly speaking) represent about 1 per cent of the skeletal weight, a result that supports the estimate proposed by Cohen and McConnaughey (2003).

This hydrated organization of the organic material must be compared with analytical results (chromatography of electrophoresis) that have more precisely established

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

the proteoglycan composition of the skeletal organic phases. 2D electrophoresis is specifically relevant (Dauphin, 2001) because Alcian blue staining allows acidic sulfated glucidic compounds to be evidenced (Fig. 12a) whereas associated proteins are silver stained (Fig. 12b). By their biochemical composition, mineralizing matrices of fibre growth layers belong to this category of glycoconjugates (proteoglycans close to aggrecan, versican, etc.), organic compounds well known for their ability to include water in their complex molecular assemblages. They usually include a core protein (MW 200 to 500 kD) with carbohydrate lateral chains, resulting in MW that may reach several megaDaltons. These data are consistent with the results concerning global characterization of organic compounds extracted from coral fibres.

A remark must be made that a 2.5–3% ratio “in weight” for hydrated organic compounds suggests that the hydrated-organic phase in coral fibres may be in a ratio about three times higher when considering their “volumes” (aragonite density: 2.99). Even assuming a high density for this hydrated organic phase (i.e. 1.5), this may result in a global proportion of about 5–7% of the skeleton volume occupied by the non-carbonate component, a result consistent with the proportion of nanograins and organic phases on the AFM pictures.

Profiles of thermal decompositions observed on the TGA experiments show that signals corresponding to the decomposition of organic compounds are observed up to 550°C (Cuif et al., 2004). This unexpected resistance of organic material to heating is explainable by the nanometric interfingering of mineral and organic components. From a practical standpoint, this raises questions about isotope ratio measurements. The heating phase included in the preparative process, before acidic decomposition of calcareous materials, is usually limited to 350/400°C, a temperature that is unable to completely remove the intra crystalline organic phases: the main part of the organic component is decomposed above this temperature. Consequently, isotopical peculiarities of the remaining organic compounds may be suspected to influence the measured ratio. Such a possibility is still more probable when using SIMS methods that do not include a thermal degradation step.

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

4.2. “The hypothesized organic matrix mediation of crystal growth (Johnston, 1977; Jackson and Gladfelter, 1985) is difficult to support”

This statement (Constantz, 1986, p. 155) illustrates the Le Tissier’s comment (1991) about the diversity of biomineralization models for coral fibres, due to the difficulty to
5 characterize organic compounds within calcareous skeletons. During decades, we were unable to connect microstructural evidences of the layered organization of fibrous or prismatic microstructural units (Cuif et al., 1981, 1997, 1998) to chemical of biochemical data obtained at a corresponding scale. For instance, information about sulfur in biominerals has long been collected (Cuif et al., 1986), but the chemical status
10 of this element was not established, no more than its close relationship with the growth steps of biominerals.

Data reported here favours the interpretation of coral fibre growth as a biologically controlled process.

4.3. A polycyclic model for the growth of the coral fibres

15 The reported structural and chemical data can be summarized in a simple model of the growth layer formation (Fig. 13).

At the starting point of a biomineralization cycle (Fig. 13a), the basal ectoderm of the polyp is in close contact with the upper surface of fibres (F1 to F3), each of them with its c-axis in global conformity with fibre length (note that orientations of the two other
20 axes differ from fibre to fibre, see Fig. 2).

4.3.1. The matrix secretion step

Although physiology of the matrix secretion is thoroughly studied (see Tambutté, 1996; Allemand, 2004), molecular organization outside the ectodermal layer is still poorly understood. Chromatographic and electrophoretic characterizations have emphasized
25 the presence of acidic high molecular weight compounds in the organic part of coral

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

skeletons (Dauphin, 2001; Dauphin and Cuif, 1997), but recent results (Allemand et al., priv. com) have also evidenced an important proportion of very low weight molecules (peptides). Owing to the importance of polysaccharides in skeletal matrices, this contrast suggests that a self-assembly process may occur within the mineralizing compartment, resulting in a polymeric organic framework. The historical observation by Goreau (1956) of a glucidic layer on the external side of the basal ectoderm of the polyp may correspond to this secretory step.

This self-assembly step may be also the explanation for the crystallographic continuity between the successive growth layers of fibres. At the beginning of the biomineralization cycle, the organic compounds involved in the hypothesized self-assembly process are in contact with the upper surface of the aragonite fibres, that are facing the basal ectoderm. Each fibre provides a template that allows the organic framework resulting from the self-assembly process to be oriented in conformity with the characteristics of each of the underlying fibres (Fig. 13b).

4.3.2. The growth layer crystallization step

To create Ca-carbonate nanograins, calcium is transported to the subectodermal space (Tambutté et al., 1995; Marshall, 1996). Many analyses have shown that both glucidic and protein matrix components have compositional peculiarities that favour interaction with mineral ions. Glucids are sulfated and very acidic in composition (Dauphin, 2001), as well as aspartic/glutamic rich proteins (Mitterer, 1978; Constantz and Weiner, 1988). Taking into account the very low pl of matrix components, the free mineral ions certainly found numerous sites to be fixed on the organic framework, starting the phase 2 of the biomineralization cycle, the crystallization process properly speaking.

In this mineralization step, the hypothesized polymeric pre-oriented organic framework may play a major role, being responsible for the position of the initial crystallization sites. Thus, after having been oriented by the underlying mineral surface, the organic framework itself may ensure the crystallographic coherence of the nanograins, an essential condition to maintain the global crystallinity of fibres. It must be noticed,

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

however, that a close SEM observation of fibres (Fig. 14a) shows that they are not so perfectly monocrystalline than believed by Bryan and Hill (1941) based on optical observations.

Within the mineralizing layer, growth of nanograins progressively reduces the space available for the initial hydrated organic medium secreted in phase 1 of the biomineralization cycle. At the end of this crystallization step (Fig. 13c), the organic phase simply appears as a cortex surrounding nanograins (see Figs. 10a and 10b).

4.3.3. The coral fibre: a matrix mediated biocrystal

In contrast to the still widely admitted monocrystalline concept, coral fibres belong to this category of matrix mediated biominerals built by repeatedly produced micron-thick growth layers. A comparison can be made with the most classical of them, the calcitic prism of *Pinna nobilis*, a Pteriomorphid Pelecypod. The following points emphasize the similarity of the growth process between mollusc prisms and coral fibres (Fig. 14).

- The micronic stepping growth mode (now evidenced also in fossil corals, Stolarski, 2003) is common to both microstructures, with quite comparable patterns. Mean thickness and coordination pattern of growth units show a global scale growth control.
- In both cases growth layers are built by nanograins.
- From a biochemical stand point, hydrated glycoconjugate components are present at the nanometric level in both structures and exhibit the same spatial relationships (for XANES pictures of *Pinna* prisms see Dauphin et al., 2003).

Not only the polycyclic mineralization process and growth mode of coral fibre agree with the reported results, but they could help to interpret some recent chemical or isotopical data that begin to raise question about the common views about environment recording by corals.

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

4.4. Relevance of the polycyclic model of coral fibre to recent results of fine scale chemical and isotopical measurements

During recent years, numerous evidences have shown that proxies deriving from mineralized skeletons do not give us a perfectly reliable information. Not only various discrepancies have been reported between environmental conditions and the values measured on biocarbonates (Allison et al., 2001; Finch et al., 2003), but when measurements are made using high resolution devices, chemical variability within a given specimen appears unexpectedly high, strongly overstepping what the influence of some physiological process (i.e. photosynthesis in reef corals) is able to explain (Rollion et al., 2003). Moreover, these unexpected variations also occur within deep sea corals (Blamart, 2002; Adkins et al., 2003). Clearly, application of simple thermodynamical laws cannot provide us with an accurate interpretation of environmental signals recorded in biominerals (Juillet-Leclerc, 2004)), and a new strategy (Lough, 2003) is now required. Data reported above concerning the fine structure and the biomineralization patterns in coral skeletons may help to explain the present difficulties.

To illustrate the rapid compositional changes that may occur in successive growth layers of polycyclic biominerals, comparizon with the Pteriomorphid Pelecypod can be continued. In the *Pinna* prisms, microprobe maps of Mg and S (Figs. 13f and 13g) exemplify the rapid compositional changes in superimposed growth layers. Obviously, the chemical layering strictly corresponds to microstructural pattern and variability along the growth process is clearly different for the two mapped elements: S varies independently of Mg.

Such a compositional diversity shows that the biomineralization layer is the basic environment recording unit and demonstrates that biological regulation does not prevent various influences to be recorded in the composition of growth layers. Although no equivalent document exists for coral skeletons, experiments have shown that mineralizing activity is very sensible to physico-chemical conditions such as pH values (Marubini et al., 2002).

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

The rapid changes in fractionation ratios evidenced by high resolution SIMS measurements in corals (Meibom, 2003; Rollion et al., 2003) are probably related to this high sensitivity, that causes surprisingly rapid variations in environmental signals. As measurements made by most of sampling methods (even by computer driven hole
5 driller, laser or ICPMS) involve several growth layers (i.e. environment recording units) variations within individual growth layers are probably higher than shown by recent papers. Only NanoSIMS method approaches the relevant measurement level.

Interestingly, the first mapping experiment applying NanoSIMS method to coral fibrous tissues (Meibom, 2004¹) has produced a result that gives the final touch to the
10 hypothesis of a biological control on the crystallization process in corals. A clear Mg signal has been observed at the top of each fibre growth layer, the role of which could be to repress the crystallization at the end of the biomineralization cycle (Fig. 13c). There is a striking contrast between the meaningless low concentration of Mg in coral fibrous aragonite measured by low resolution method and the biologically significant
15 interpretation that can be proposed when Mg localized distribution is evidenced at the relevant level.

Clearly, to develop the “new strategy” that must be now elaborated to improve the use of coral proxies (Lough, 2003), a detailed understanding of biomineralization process is a prerequisite, an approach that should be based on the notion of “environment record-
20 ing unit”. Precise relationships between environmental conditions and the composition of a given biomineral layer have to be established and reciprocally, signals created by the recording process within each skeleton growth layer have to be measured at the relevant scale.

¹Meibom, A., Cuif, J. P., Hillion, F.; Constantz, B. R., Juillet-Leclerc, A., Dauphin, Y., Watanabe, T., and Dunbar, R. B.: Distribution of magnesium in coral skeleton, *Geophys. Res. Lett.*, submitted, 2004

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[I◀](#)[▶I](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Print Version](#)[Interactive Discussion](#)

5. Conclusions

1. Coral fibres are built by superimposition of few micron-thick growth layers.
2. Organic compounds are distributed within each growth layer at a sub-micronic scale.
3. Growth layers are made of mineral nanograins densely packed within an organic phase.
4. Concentric growth patterns within a given corallite indicate that the growth process is coordinated by polyp physiology.
5. Consequently, each growth layer is the basic *Environment Recording Unit*.

References

- Adkins, J. F., Boyle E. A., Curry, W. B., and Lutringer A.: Stable isotopes in deep-sea corals and a new mechanism for “vital effects, *Ceochimica et Cosmochimica Acta*, 67, 6, 1129–1143, 2003.
- Al-Horani, F. A., Al-Moghrabi, S. M., and de Beer, D.: The mechanism of calcification and its relation to photosynthesis and respiration in the scleractinian coral, *Galaxea fascicularis*, *Marine Biology*, 142, 419–426, 2003.
- Allemand, D., Ferrier-Pages, C., Furla, P., Houlbèque F., Puverel S., Reynaud S., Tambutté E., Tambutté S., and Zoccola, D.: Biomineralization in reef-building corals: from molecular mechanisms to environmental control. *C.R. Acad. Sc; Paris*, in press, 2004.
- Barnes, D. J.: Coral skeletons: An explanation of their growth and structure. *Science*, 170, 1305–1308, 1970.
- Blamart, D., Cuif, J. P., and Juillet-Leclerc, A.: O-stable isotopes distribution in deep-sea corals from SIMS measurements, 27th Gen. Ass. European Geophysical Soc., *Geophys. Res. Abstr.*, 4, EGS02-A-01713, Nice 21–26 April, 2002.
- Bryan, W. H. and Hill, D.: Spherulitic crystallization as a mechanism of skeletal growth in the hexacorals, *Proc. Roy. Soc. Queensland*, 52/9, 78–91, 1941.

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

- Cohen, A. L. and McConnaughey, T. A.: Geochemical perspective on coral mineralization, *Rev. Mineral. Geochem.*, 54, 151–187, 2003.
- Constantz, B.: Coral Skeleton Construction: a Physiochemically dominated Process, *Palaios*, 1, 152–157, 1986.
- 5 Constantz, B. and Weiner, S.: Acidic macromolecules associated with the mineral phase of scleractinian coral skeletons, *J. Exp. Zool.*, 248, 253–58, 1988.
- Cuif, J. P., Denis, A., and Gaspard, D.: Recherche d'une méthode d'analyse ultrastructurale des tests carbonatés d'Invertébrés, *Bull. Soc. géol. Fr., sér.*, 9, 28/5, 525–534, 1981.
- Cuif, J. P., Dauphin, Y., Flamand, D., Frerotte, B., and Gautret, P.: La mesure localisée du taux de soufre comme indicateur de l'origine et de l'état diagénétique des biocristaux carbonatés
- 10 *C. R. Acad. Sc. Paris*, 303, II, 3, 251–256, 1986.
- Cuif, J. P., Dauphin, Y., Denis, A., Gautret, P., Kiyashko, S., and Massault M.: Facteurs de la diagenèse précoce des biominéraux: exemple d'un polypier de Porites de Nouvelle Calédonie, *Geobios*, MS 20, 171–179, 1997.
- 15 Cuif, J. P., Dauphin, Y., and Gautret, P.: Biomineralization features in scleractinian coral skeletons: source of new taxonomic criteria, *Bol. Real. Soc. Hist. Nat. (Geol.)*, 92, 1–4, 129–141, 1997.
- Cuif, J. P. and Dauphin, Y.: Microstructural and physico-chemical characterization of “centres of calcification” in septa of some recent Scleractinian corals, *Pal. Zeit.*, 72, 3/4, 257–270, 1998.
- 20 Cuif, J. P., Dauphin, Y., and Gautret, P.: Compositional diversity of soluble mineralizing matrices in some recent coral skeletons compared to fine-scale growth structures of fibres. Discussion of consequences for biomineralization and diagenesis, *Int. Journ. Earth Sciences*, 88, 582–592, 1999.
- Cuif, J. P., Dauphin, Y., Berthet, P., and Jegoudez, J.: Associated water and organic compounds in coral skeletons: quantitative thermogravimetry coupled to infrared absorption spectrometry, *Geochemistry, Geophysics, Geosystems*, in press, 2004.
- 25 Dauphin, Y.: Comparative studies of skeletal soluble matrices from some Scleractinian corals and Molluscs, *Int. Journ. of Biol. Macromol.*, 28, 293–304, 2001.
- Dauphin, Y. and Cuif, J. P.: Isoelectric properties of the soluble matrices in relation to the chemical composition of some Scleractinian skeletons, *Electrophoresis*, 18, 1180–1183, 1997.
- 30 Dauphin, Y., Cuif, J. P., Doucet, J., Salomé, M., Susini, J., and Williams, C. T.: In situ mapping of growth lines in the calcitic prismatic layers of molluscs shells using X-ray absorption near edge structure (XANES) spectroscopy at the sulphur k-edge, *Marine Biology*, 142, 299–304,

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

2003.

- Gaffey, S.: Water in skeletal carbonates, *J. Sedimentary Petrology*, 58/3, 397–414, 1988.
- Gautret, P., Cuif, J. P., and Freiwald, A.: Composition of soluble mineralizing matrices in Zooxanthellate and Non-Zooxanthellate Scleractinian corals: biochemical assessment of photosynthetic metabolism through the study of skeletal features, *Facies*, 36, 189–194, 1997.
- 5 Goreau, T.: Histochemistry of mucopolysaccharide – like substances and alkaline phosphatase in Madreporaria, *Nature*, 177, 1029–1030, 1956.
- Goreau, T. F.: The physiology of skeleton formation in corals. I. A method for measuring the rate of calcium deposition by corals under different conditions, *Biol. Bull.*, 116, 59–75, 1959.
- 10 Heider, A. von: Die Gattung *Cladocora* Ehrenb. Sitzungber. d. K. Akad. d. Wiss. Wien, 44, 707–735, 1881.
- Hidaka, M.: Surface structure of skeletons of the coral *Galaxea fascicularis* formed under different light conditions, *Proc. 6th Int Coral reef Symp.*, 3, 95–100, 1988.
- Hidaka, M.: Deposition of fusiform crystals without apparent diurnal rhythm at the growing edge of septa of the coral *Galaxea fascicularis* Coral Reefs, 10, 41–45, 1991.
- 15 Johnston, I. S.: Aspect of a skeletal organic matrix and the process of skeletogenesis in the reef coral *Pocillopora damicornis*, *Proc. third Int. Coral Reef Cong.*, Brisbane, 2, 301–320, 1977.
- Johnston, I. S.: The ultrastructure of skeletogenesis in hermatypic corals. *Internat. Review of Cytology*, 67, 171–213, 1980.
- 20 Koch, G. von: Ueber die Entwicklung des Kalkskeletes von *Astroides calycularis* u. dessen Morphologische Bedeutung, *Mitth. Zool. St. Neapel*, 3, 284–292, 1882.
- Le Tissier, M. D'A. A.: The nature of the skeleton and skeletogenetic tissues in the Cnidaria, *Hydrobiologia*, 216/217, 397–402, 1991.
- 25 Lough, J. M.: A strategy to improve the contribution of coral data to high-resolution paleoclimatology, *Paleoceanography, Paleoclimatology, Paleontology*, 204, 115–143, 2003.
- Marshall, A. T.: Calcification in hermatypic and ahermatypic corals, *Sciences*, 271, 637–639, 1966.
- Marubini, F., Ferrier-Pages, C., and Cuif, J. P.: Suppression of skeletal growth in scleractinian corals by decreasing ambient carbonate-ion concentration: a cross-family comparison, *Proc. R. Soc. Lond.*, B 270, 179–184, 2002.
- 30 Mitterer, R. M.: Amino acid composition and metal binding capability of the skeletal protein of corals, *Bull. Mar. Sci.*, 28/1, 173–180, 1978.

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

- Stolarski, J.: Three-dimensional micro- and nanostructural characteristics of the scleractinian coral skeleton: A biocalcification proxy, *Acta. Paleont. Pol.*, 48, 4, 497–530, 2003.
- Tambutté, E., Allemand, D., Bourge, I., and Jaubert J.: An improved Ca45 protocol for investigating physiological mechanisms in coral calcification, *Mar. Biol.*, 122, 453–459, 1995.
- 5 Tambutté, E.: Processus de calcification d'un Scléractiniaire hermatypique, *Stylophora pistillata* (Esper, 1797), Thèse Univ. Nice, 247 p., 1996.
- Wada, K. and Fujinuki, T.: Physiological regulation of shell formation in Molluscs, I. Chemical composition of extrapallial fluids, *Bull. Natl. Pearl Res. Lab.*, 18, 2085–2110, 1974.
- Wada, K. and Fujinuki, T.: Biomineralization in bivalve molluscs with emphasis on the chemical
 10 composition of the extrapallial fluid, *The Mechanisms of Mineralization in the Invertebrates and Plants*, edited by Watabe, N. and Wilbur, K. M., The Belle W. Baruch library in Marine Science, 5, 175–190, 1976.
- Wainwright, S. A.: Skeletal organization in the coral *Pocillopora damicornis*, *Quart. J. micr. Sci.*, 104/2, 169–183, 1963.
- 15 Wilfert, M. and Peters, W.: Vorkommen von Chitin bei Coelenteraten, *Z. Morph. Tiere*, 64, 77–84, 1969.
- Young, S. D.: Organic materials from scleractinian coral skeletons, I Variation in composition between several species, *Comp. Biochem. Physiol.*, 40B, 113–120, 1971.

BGD

1, 625–658, 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

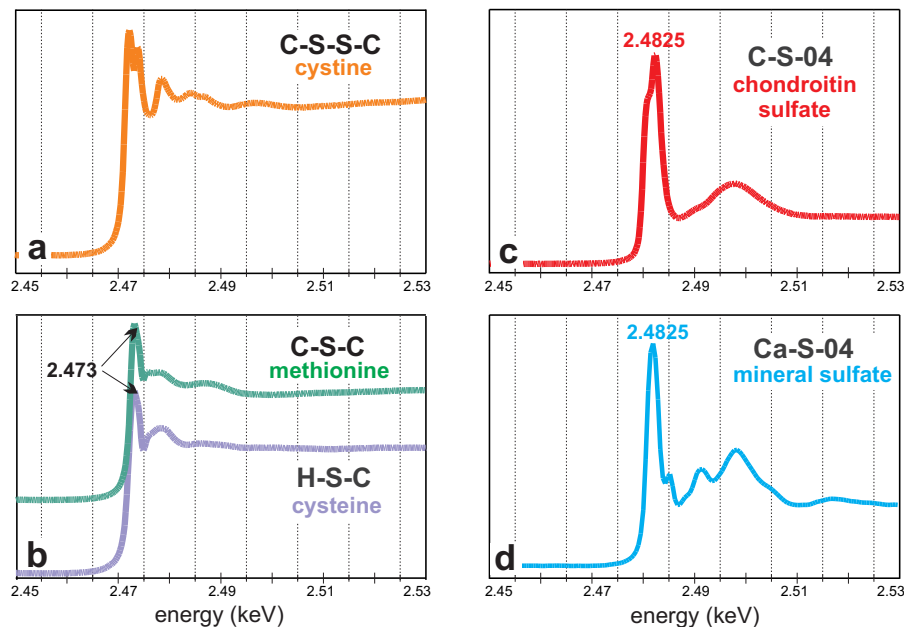


Fig. 1. Differences between XANES spectra from four sulfur organic bounds and sulfur in a calcium sulfate. Not only is the edge energy significant, but also the “near edge” oscillation: in the SO_4 case, it allows to clearly separate the sulfur linked to an organic carbon from the sulfur in the mineral species.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

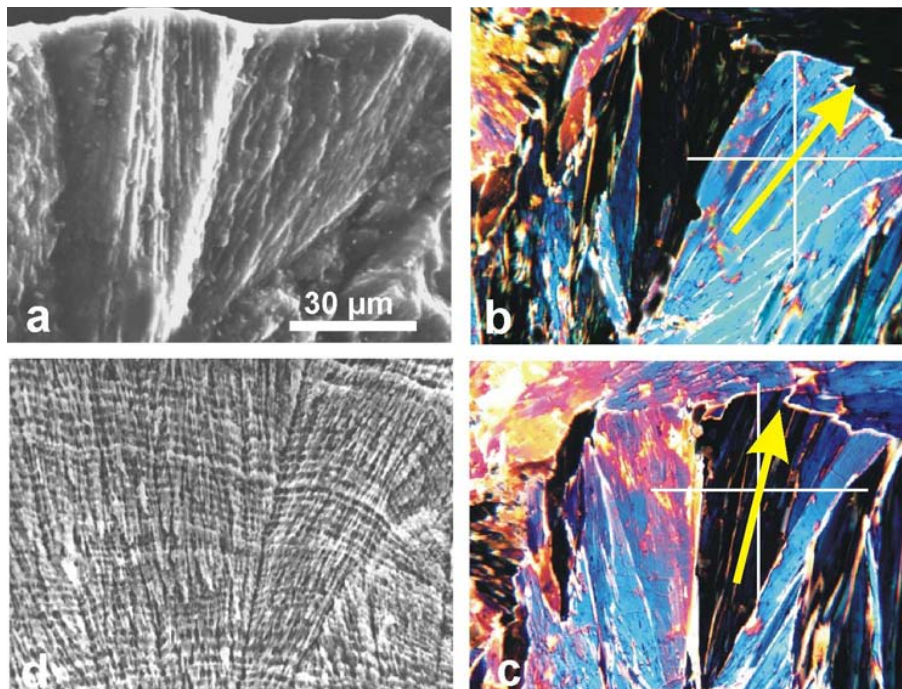


Fig. 2. Three different aspects of the same group of coral fibres. **(a):** Morphology of fibres on a fracture surface of a corallite (*Favia stelligera*). **(b)–(c):** Ultra-thin section observed in polarized light. Fibres are groups of subunits with globally similar behaviour with respect to polarization directions (orientation of the yellow arrow). **(d)** Aspect of the same skeletal sector after polishing and enzymatic etching. Global organization of fibres is still recognizable, but growth layers have been made visible by synchronous differences in sensitivity to etching. This results in coordinated changes in thickness and spacing of growth layers, allowing a precise description of fibre growth to be made.

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

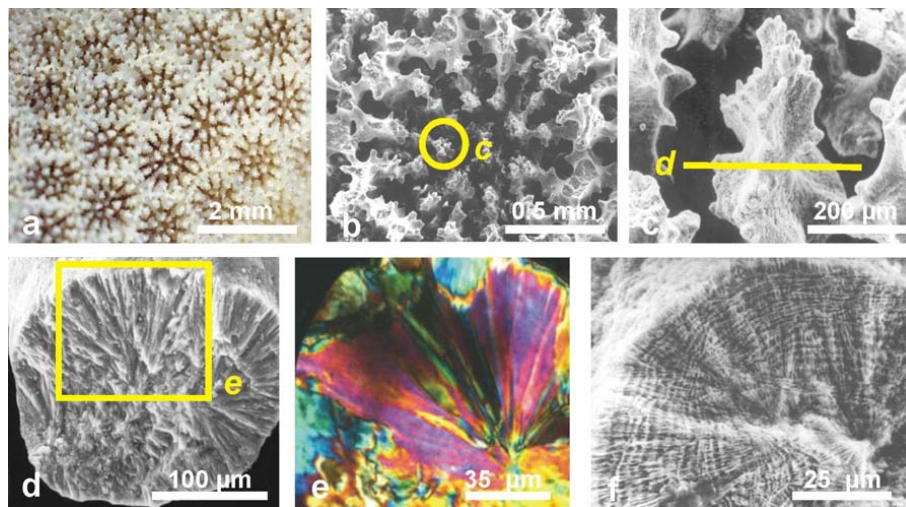


Fig. 3. From morphology to elemental growth layer in a *Porites* skeleton. **(a)–(b):** Morphology of *Porites* *c.f.* *australiensis* corallite. **(c)** Longitudinal view of a vertical unit. **(d)** Radial disposition of fibres in a vertical unit. **(e)** Ultra-thin slide in a vertical unit: radial disposition of fibre fans is well visible, a typical example of what led numerous authors to emphasize the similarity of coral fibres with abiotic crystallisations. **(f)** Etching of the fractured surface in the *Porites* corallite, evidencing the fibre growth layers.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

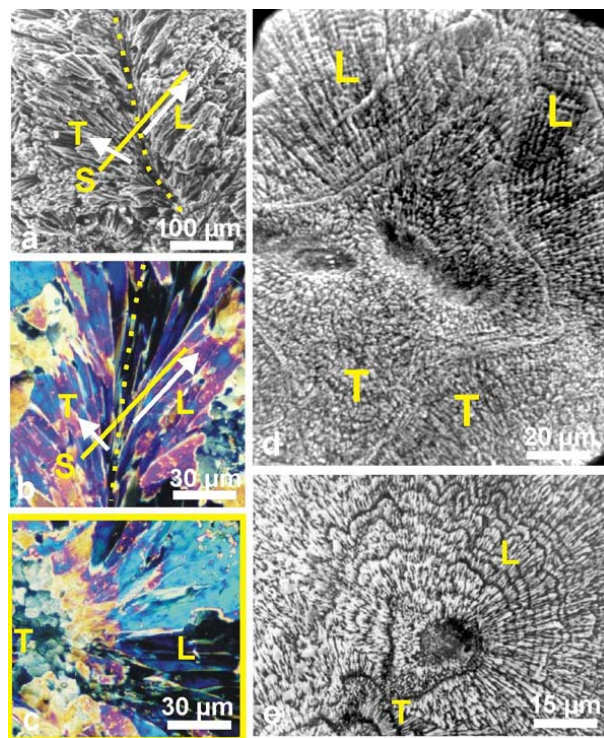


Fig. 4. Diversity of fibre directions on a corallite section, and consequences on the observation of etching results. **(a)** Symmetrically diverging fibres on both sides of a septum median plan (yellow dotted line). **(b)** Diverging fibres observed in thin slides, polarized light. S is the surface transverse to the view. Depending on their orientation, fibres may appear transversely (T) or longitudinally (L) cut. **(c)** This yellow framed view corresponds to the b transverse section. On the left side, fibres appear as polygonal units because they are transversely cut (T). On the opposite they are longitudinally cut (L). **(d)–(e)** When looking at etched surfaces, fibre growth layers are well visible in fibre longitudinal sections only: d: *Leptoria phrygia*; e: *Diploastrea heliopora*.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

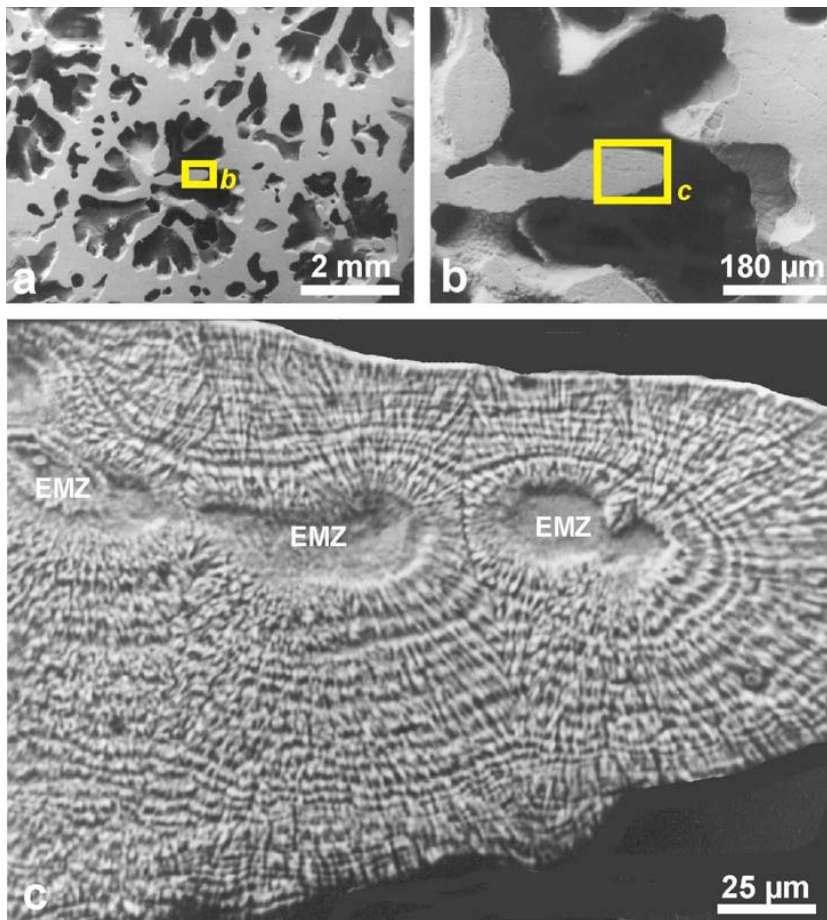


Fig. 5. Synchronism of growth layers in a septum of *Favia stelligera*. Concentric growth layering indicates that mineralizing activity of the ectodermal cell layer is controlled at a global level, in contrast to the usual concept of “crystal growth competition”.

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

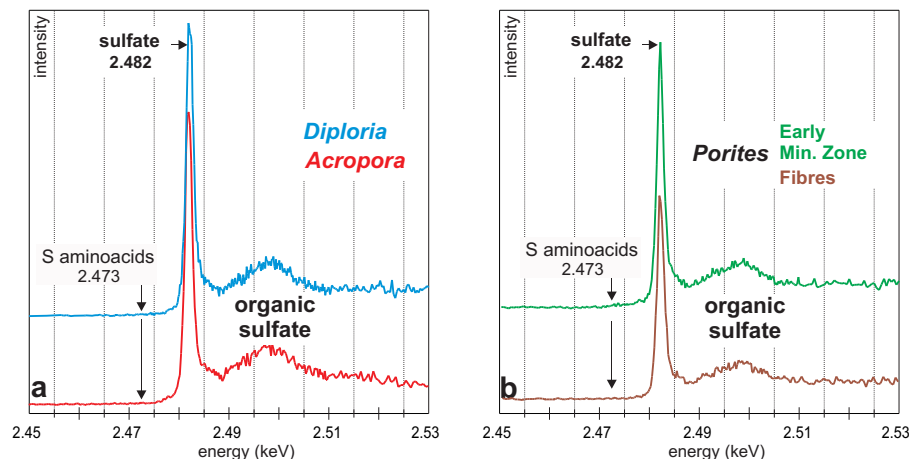


Fig. 6. Responses of polished surfaces in corallites of *Diploria labyrinthica*, *Acropora digitiformis* and *Porites* c.f. *australiensis* to a 2.4825 keV X-ray beam. In all cases a strong signal corresponding to a sulfur bond in organic sulfates is detected, whatever the location of the X-ray beam on the corallite surface, including the Early Mineralization Zones (the “centres of calcification”). In contrast, no response is obtained for S-bond in amino acids.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

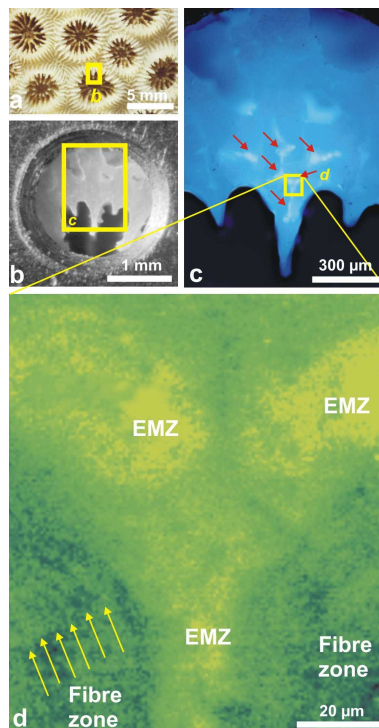


Fig. 7. Mapping of sulfated polysaccharides in *Montastrea curta*. **(a)** Morphology of the coral-lites. **(b)** Specimen submitted to analyse, in the sample holder. **(c)** Selection of the region to be mapped by UV fluorescence: the Early Mineralization zones show a strong response (red arrows). **(d)** Biochemical map of the selected zone. The Early Mineralization Zones exhibit a high sulfated polysaccharide concentration. In fibres, the well visible banding pattern in exact conformity with fibre growth layers (arrows) shows that mineral phase and sulfated polysaccharides are associated at a submicronic scale within fibres.

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

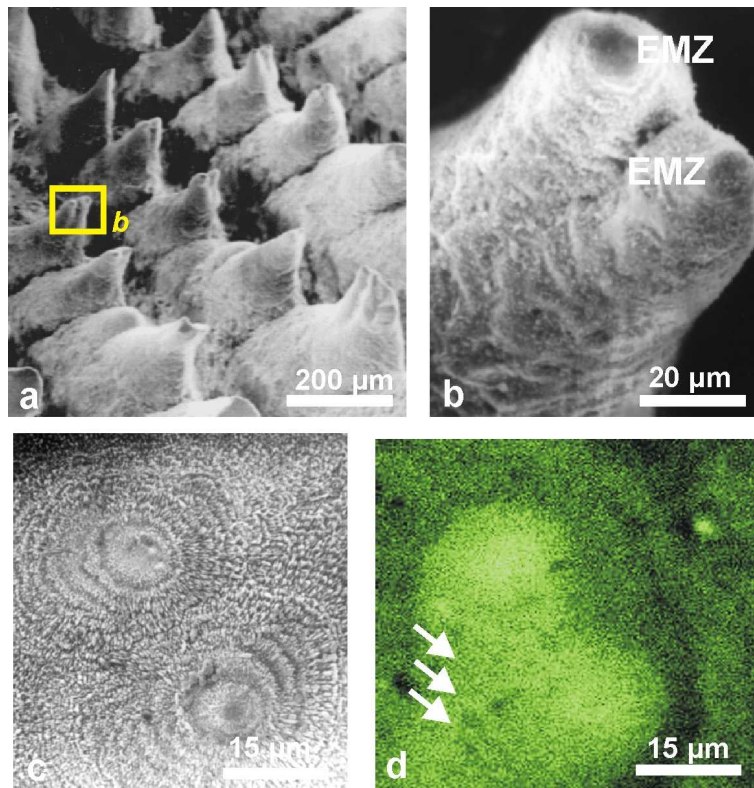


Fig. 8. Growth layers and sulfated polysaccharide repartition in *Diploastrea labyrinthica*. **(a)** Morphology of growing edges of septa built by a simple series of conical units. **(b)** At the tip of conical units, the Early Mineralization Zone (EMZ) surrounded by the beginning of fibrous zone **(c)** Etched section, showing the concentric growth layers surrounding the EMZ. **(d)** XANES mapping of organic sulfur provides a very comparable pattern.

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

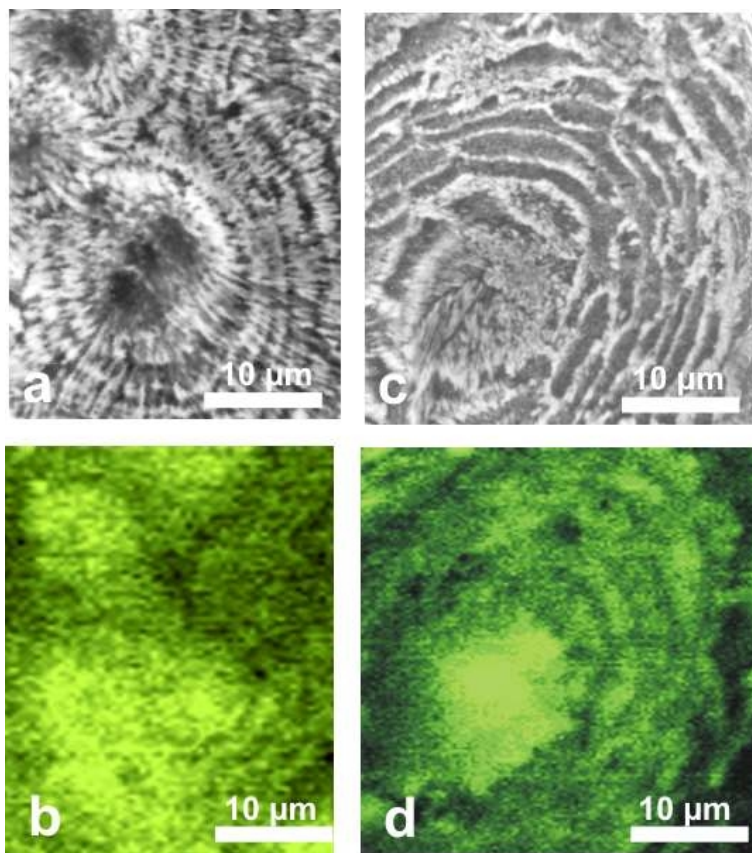


Fig. 9. Microstructural patterns and corresponding XANES mapping of organic sulfur bond on polished surfaces in *Porites c.f. australiensis* (a)–(b) and *Acropora digitifera* (c)–(d).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

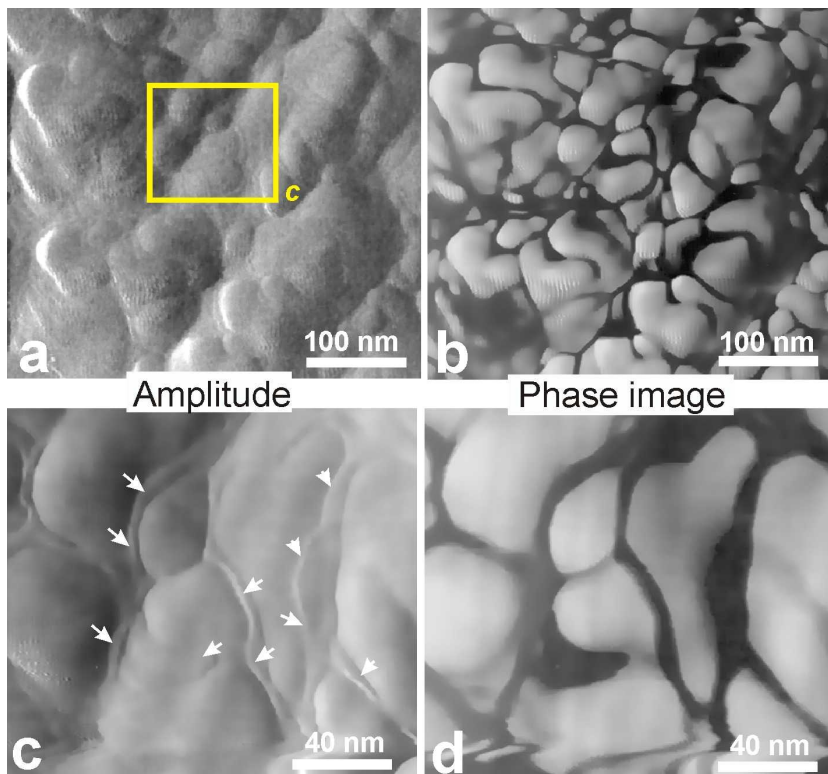


Fig. 10. AFM images of skeletal nanograins in *Merulina scabricula*. Amplitude images (**a**) and (**c**) and phase images (**b**) and (**d**) show the dual composition of skeletal grains. Phase imaging reveals the importance of the very weak relief that can be seen on a and c pictures of the grain surfaces (arrows). The very high contrast produced by these structures demonstrates that they are basically different, from a chemical standpoint, from the nanograins themselves. The XANES in situ characterization of sulfated polysaccharides at a submicronic scale fully supports the interpretation of this high phase-contrast material as an organic coating of skeletal grains.

**The environment
recording unit in
coral skeletons**

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

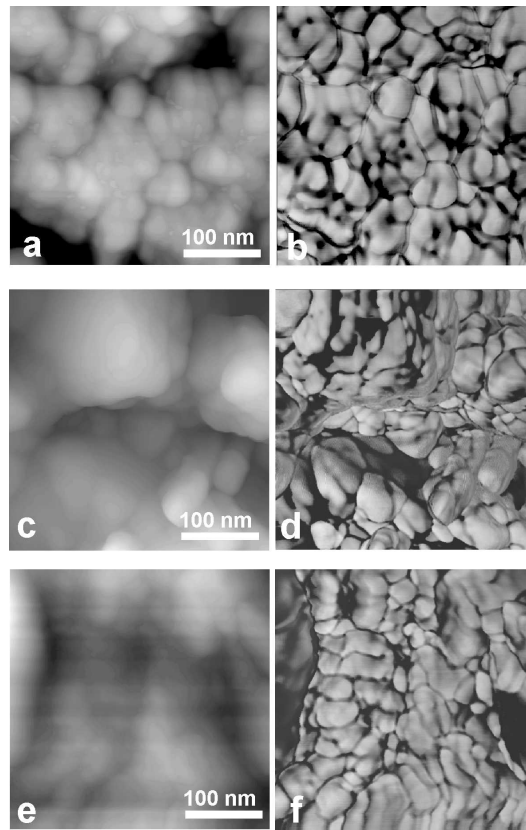


Fig. 11. Equivalent AFM amplitude and phase pictures from corallite belonging to three species: **(a)–(b)**: *Favia stelligera*; **(c)–(d)**: *Cladocora caespitosa*; **(e)–(f)**: *Caryophyllia am-brosia*. Note that surface topography remains rather unprecise on height images (a, c, e), but phase imaging (b, d, f) provides clear information about the basic granular structure, that has been found in all observed coral skeletons.

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

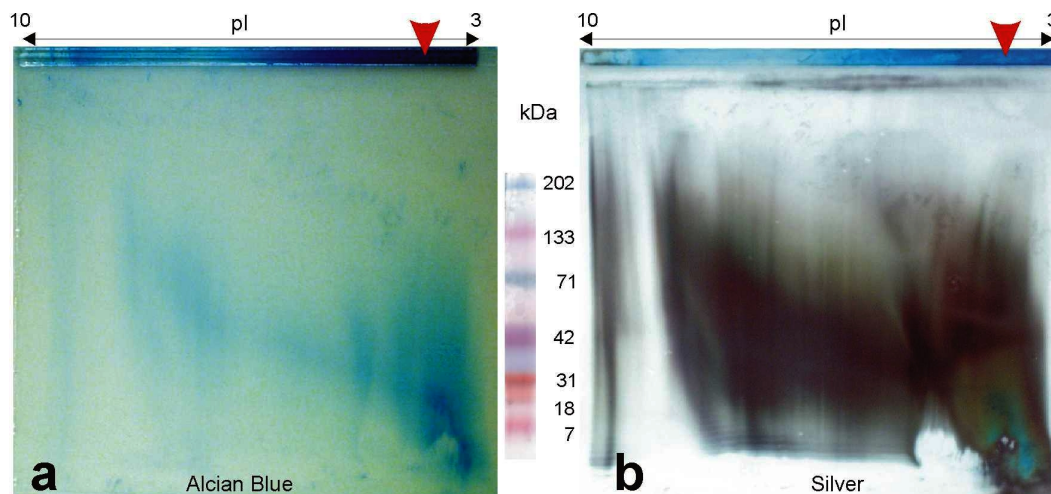


Fig. 12. 2D electrophoretic characterization of sulfated polysaccharides **(a)** and protein compounds **(b)** from coral skeletons. Alcian blue staining of acidic sulfated polysaccharides shows that, after isoelectric focussing that establish their low pI, most of them remain included at the top of the 2D gel (arrows), showing that their molecular weight is higher than 300 kD, that is the upper MW accepted by the mass characterization gel.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

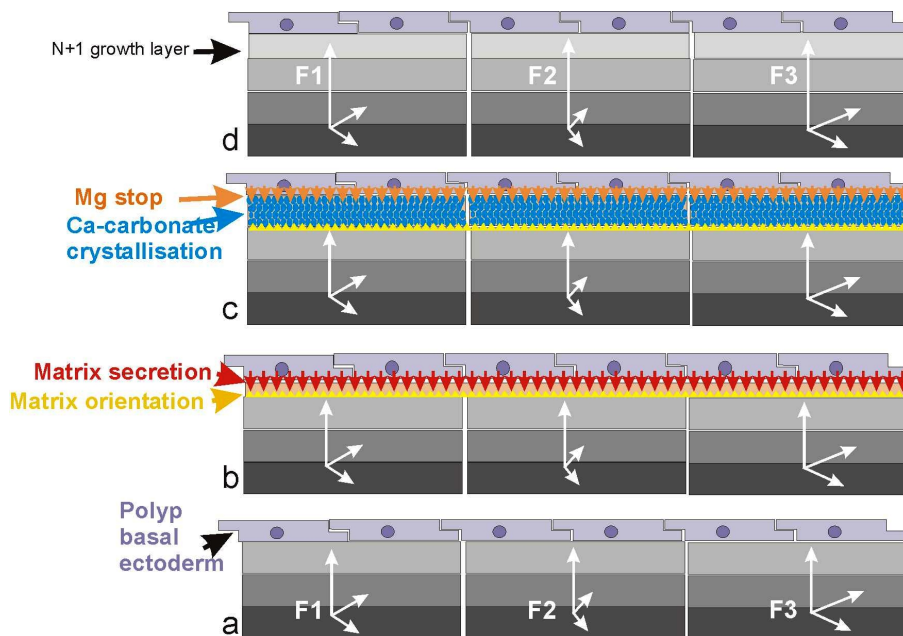


Fig. 13. Scheme of a growth layer formation in a coral skeleton, summarizing the structural, chemical and biochemical data.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004

The environment recording unit in coral skeletons

J. P. Cuif and Y. Dauphin

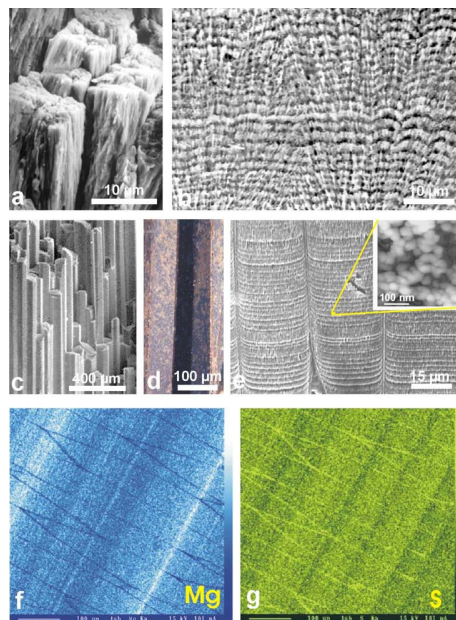


Fig. 14. Similarity between the fine structure of coral fibrous tissue and a typical “matrix mediated” mollusc microstructure: the prismatic layer of *Pinna nobilis*. **(a)** Close view of coral fibres: each fibre is not single crystal but a cluster of elongated subunits with globally similar polarizing behaviour (see Fig. 2). **(b)** Polished and etched surface in the fibrous zone of a coral skeleton: fibre subunits are still visible but the dominant pattern is the stepping growth mode. **(c)–(e)** Morphology, ultra-thin slide (polarized light) and etching surface in the *Pinna* prismatic shell layer. Calcitic prism morphology (c) is more precisely defined than coral fibres, due to their strong organic envelope, and monocrystalline behaviour (d) of the prisms is also better marked. However, an obvious similarity between the fine structures of coral fibres appears after etching (e). **(f)–(g)** Microprobe maps (Mg and S) on a longitudinal section of the *Pinna* prismatic shell layer. Note the fine layering of minor element repartition, the high contrast between successive growth layers. Also remarkable is the independence in concentration changes between the two elements (Microprobe mappings by C.T. Williams, NHM London).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Print Version

Interactive Discussion

© EGU 2004